



AAHS/ASPN/ASRM Joint Outstanding Paper Session
Saturday, January 14, 2017, 11:00am – 12:00pm

1. Trainee Selection and Correlation with Cognitive and Microsurgical Technical Skills

Department of Plastic Surgery, MD Anderson Cancer Center, Houston, 232, USA

Mark V Schaverien, MD MEd MSc FRCS; MD Anderson Cancer Center, Houston, TX; Charles E Butler, MD FACS; Plastic Surgery, University Of Texas MD Anderson Cancer Center, Houston, TX; Jun Liu, PhD; Plastic Surgery, The University of Texas MD Anderson Cancer Center, Houston, TX; Jesse C Selber, MD, MPH, FACS; Department of Plastic Surgery, MD Anderson Cancer Center, Houston, TX

BACKGROUND: The selection process for surgical trainees aims to identify those who will perform best during training and have the greatest potential as future surgeons. Better understanding the predictive relationship between interview performance, level of technical skill, and performance during training will allow optimisation of the interview and evaluation process to identify the best candidates.

METHODS: Three annual cohorts of Reconstructive Microsurgery fellows at MD Anderson Cancer Center, comprising 20 trainees, were included in the study. At interview, subjects were rated using seven criteria, as well as given a score for overall impression. At the start and end of the fellowship, microsurgical technical skill was assessed both in the laboratory and OR using a validated tool. At the end of the fellowship there was a final evaluation of performance using criteria adapted from the six Accreditation Council for Graduate Medical Education (ACGME) core competencies. Scores at interview, technical skill assessment, and final evaluation scores were all compared in multiple ways to determine associations and predictive factors.

RESULTS: Microsurgical skill assessment in the OR at the start of training correlated with all domains evaluated at interview, most closely with Plastic Surgery Training Experience. Microsurgical skill assessment in the OR at the end of training also correlated with scores on the majority of final assessment criteria based on ACGME core competencies, with the highest correlations with Patient Care and Medical Knowledge. Assessment of microsurgical skill in the laboratory at the start of the fellowship did not improve the predictive relationship between interview scores and ACGME core competency evaluations.

CONCLUSIONS: Microsurgical technical skill in the OR tracked with all domains evaluated at interview, and also with all ACGME core competency evaluations. These results validate the use of the current selection process in choosing candidates with the highest level of both cognitive

and technical skill, and also support the effectiveness of the one-year microsurgical fellowship in improving microsurgical skill in all trainees.

2. Muscle Graft Volume Implanted in Regenerative Peripheral Nerve Interfaces Influences Electrical Signal Transduction

University of Michigan, Ann Arbor, 212, USA

Yaxi Hu, MD¹; Alixandra L VanBelkum, BA²; Ian C Sando, MD¹; Racquel Sohasky, Undergraduate³; Jana D Moon, BS⁴; Stephen WP Kemp, PhD¹; Paul S Cederna, MD⁴; Melanie G Urbanchek, PhD⁴; (1)University of Michigan, Ann Arbor, MI, (2)Department of Surgery, Section of Plastic Surgery, University of Michigan, Ann Arbor, MI, (3)University of Michigan, Ann Arbor, MI, (4)Plastic Surgery, University of Michigan, Ann Arbor, MI

Introduction: Regenerative Peripheral Nerve Interfaces (RPNI) for interfacing to prostheses are formed by combining biology, biomedical abiotic products, muscle regeneration, nerve reinnervation, and tissue revascularization. An RPNI unit is a small muscle graft which is moved to the end of a transected residual nerve. The nerve fascicle is implanted / neurotized into the grafted muscle and electrodes are implanted with the muscle graft. Standard RPNI small muscle grafts have weighed approximately 150 mg with dimensions of 3 cm long by 1 cm diameter. In rat studies, RPNI maintained muscle volume and signaling capacities for at least 9 months once healing processes stabilized at 3 months post implant. Though small muscle grafts conserve skeletal muscle, there are advantages to grafting larger volumes. Our purpose was to evaluate RPNI signaling capacity with respect: a) to increasing grafted skeletal muscle volume and b) to time of implantation.

Methods: F344 male rats (n=30) were assigned to 1 of 4 groups defined by RPNI muscle graft volume: 150mg (RPNI-150), 300 mg (RPNI-300), 600 mg (RPNI-600) or 1200 mg (RPNI-1200). Each RPNI consisted of a semimembranosus allograft to the upper thigh and neurotized by the transected peroneal nerve. All grafts were 3 cm in length. Group 1-4 RPNI recovered for 3 months while a second 1200 mg group 5 (RPNI-1200yr) recovered for 1 year post-surgery. Compound muscle action potential (CMAP), RPNI contractile force, and RPNI histology were evaluated.

Results: All rats were healthy with RPNI that were well vascularized and responded to stimuli applied to the implanted peroneal nerve. Data are summarized in Table 1. At evaluation, RPNI of heavier graft volume still weighed more than the lighter RPNI; however, heavier grafts retained less of their original muscle mass (Fig 1). Signaling capacities or the magnitude of electrophysiology (CMAP) and force measurements were inversely related to the volume of the muscle graft. Extending recuperation time for large volume grafts did not improve signaling.

Conclusions: RPNI with muscle grafts weighing between 150 and 300 mg are optimal for signal production in the rat model. One year for recuperation did not benefit RPNI implanted with heavier grafts.

Table 1: Summary for RPNI of four volumes and two time points and ANOVA results

| | RPNI SURGICAL GROUPS | | | | |
|-----------------------------------|----------------------|-------------|-------------|--------------|-------------|
| | RPNI-150 | RPNI-300 | RPNI-600 | RPNI-1200 | RPNI-1200yr |
| Muscle mass (mg) implanted | 171±16 | 329±16 | 614±24 | 1211±48 | 1056±294 |
| Number of rats | 6/6 | 6/6 | 5/6 | 4/6 | 4/4 |
| Time implanted | 3 months | 3 months | 3 months | 3 months | 1 year |
| Body mass (g); end | 385±14 | 381±14 | 386±11 | 379±6.8 | 438±8.6* |
| Muscle mass (mg) end | 72±7 | 98±4 | 140±11 | 184±36** | 275±114* |
| Muscle mass retained | 41% | 29%** | 23%** | 15%*** | 25%** |
| CSA (mm²) end | 7.04±1.9 | 9.54±0.71** | 13.31±1.7** | 13.34±2.52** | 16.05±3.19* |
| CMAP (mVms) | 5.4±2.02 | 4.6±2.5 | 3.4±1.3 | 2.9±1.7 | 1.6±0.74*** |
| Amplitude (mV) | 6.6±3.2 | 4.7±1.9 | 3.1±1.3 | 2.7±1.5 | 1.4±0.73*** |
| Max Force (mN) | 289±106 | 259±122** | 235±169** | 91±43**** | 117±118** |

Values are mean ± 1 standard deviation. Abbreviations: imp, implanted; end, at final evaluation; g, grams; mg, milligrams; CSA, muscle cross sectional area; *different from RPNI-150, RPNI-300, and RPNI-600; **different from RPNI-150; *** different from RPNI150 and RPNI-300, **** different from all other groups. Level of significance was p<0.05.

Images were captured with a 4x objective

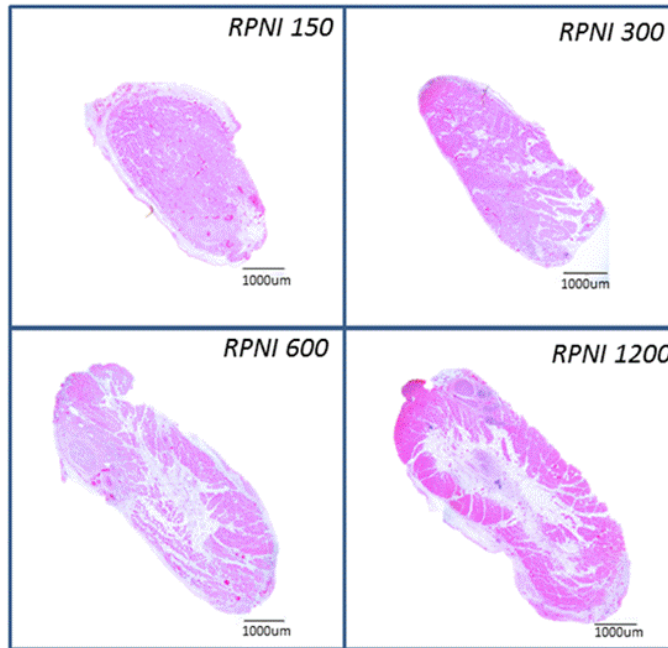


Figure 1. Representative H&E stained cross sections from RPNI muscles implanted for 3 months. Note the lack of muscle fiber regeneration in the centers of the RPNI-600 and RPNI-1200.